



Studies towards the total synthesis of novel dolabellane-type diterpenoids: construction of the 5,11-fused bicyclic framework

Goverdhan Mehta* and Jayant D. Umarye

Department of Organic Chemistry, Indian Institute of Science, Bangalore 560 012, India

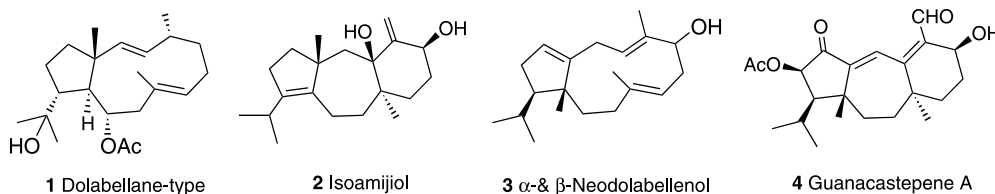
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Abstract—An oxy-Cope protocol has been grafted on to a previously crafted hydroazulenenic system to provide a facile entry into the 5,11-fused bicyclic skeleton present in the novel neodolabellane diterpenoids. Formation of a novel tricyclic diterpenoid framework, generated through a transannular cyclization in the 11-membered ring present in the neodolabellane skeleton and embodying a bridged cycloheptatriene moiety, has also been encountered.

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Dolabellane diterpenes bearing a novel 5,11-fused bicyclic framework **1** were for the first time encountered way back in 1976 from the digestive glands of the sea hare *Dolabella California* sterns^{1a} and since then natural products based on this skeleton have surfaced from marine and terrestrial sources at regular intervals, constituting one of the largest group among diterpenoids.¹ Sharing close biogenetic relationship with dolabellanes are the 5,7,6-fused tricyclic diterpenoids of dolastane-type **2** (isoamijiol)^{2c} which are also widely occurring in Nature.² Both dolabellane and dolastane diterpenoids have aroused a great deal of synthetic interest both on account of the novelty of their carbocyclic skeleta and the dense oxy-functionalization present in them.^{3,4}

reported⁶ from an unidentified endophytic fungus growing on the tree *Daphnopsis Americana* and exhibiting remarkable antibiotic activity. Both, neodolabellane and guanacastepene diterpenoids constitute challenging targets for synthesis. While several groups have been in pursuit of guanacastepenes,⁷ the group of Williams⁸ is the only one to report the synthetic accomplishment of neodolabellane natural products. In view of our ongoing interest in the synthesis of diterpenoids **1–4**,^{3o,4j,7a–c} we have devised an entry into the highly functionalized 5,11-fused bicyclic system present in neodolabellanes in which an oxy-Cope rearrangement is the pivotal step and these and related studies are reported in this letter.



Subsequently, the neodolabellane skeleton represented by **3**, α- and β-neodolabellenol,^{5c} and biogenetically derived from a dolabellane precursor through stereospecific methyl migration was also reported from marine organisms.⁵ More interestingly, a 5,7,6-fused tricyclic diterpeneoid guanacastepene A **4**, having the same biogenetic kinship with neodolabellanes, as dolastanes **2** have with dolabellane **1**, has been very recently

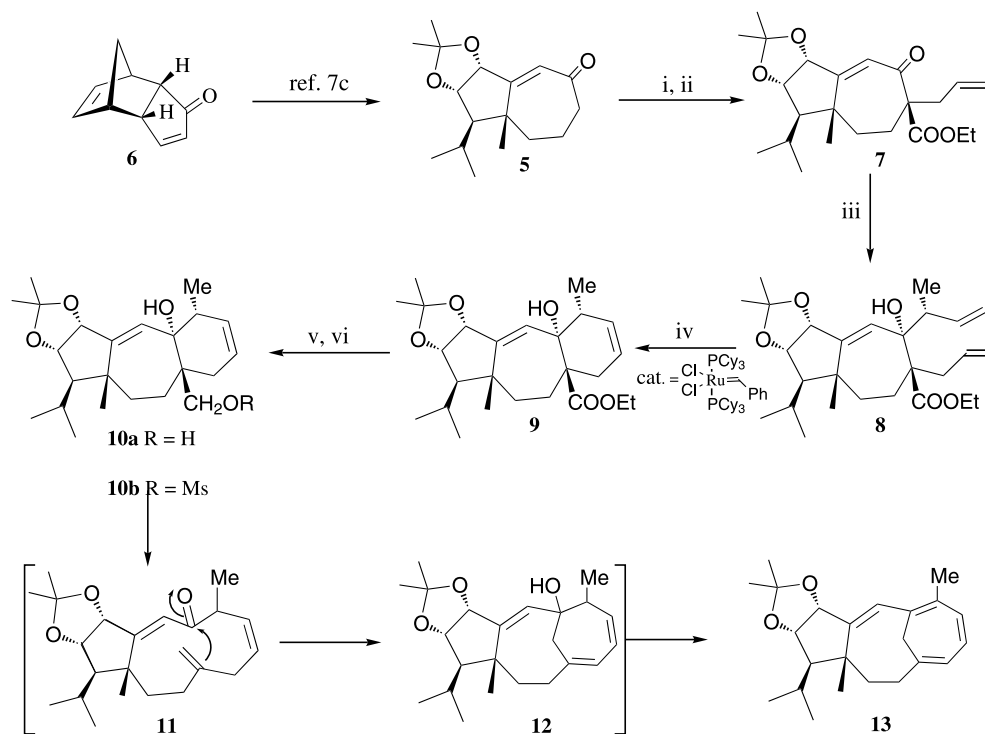
We have recently described the synthesis of hydroazulenone **5** from dicyclopentadienone **6** in the context of our approach to the synthesis of guanacastepene A **4**.^{7a–c} The seven membered ring of hydroazulenone **5**, having the angular methyl and the isopropyl groups in correct *cis*-stereochemical disposition, appeared well poised for the execution of a four carbon ring enlargement protocol to deliver the desired 5,11-fused bicyclic ring system of neodolabellanes. Our first foray in this context involved a four carbon ring annulation followed by a Grob-type fragmentation. Consequently,

* Corresponding author.

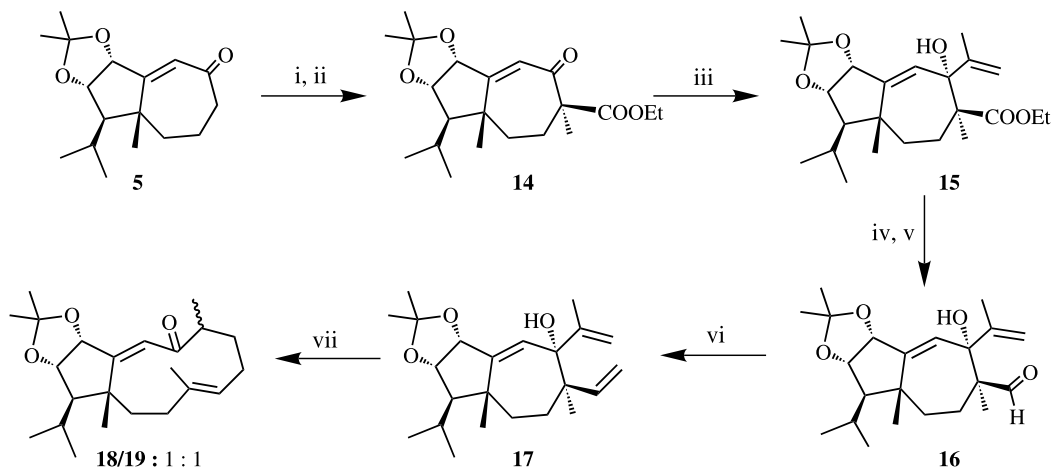
hydroazulenone **5** was elaborated to the tricyclic system **9** through a sequence involving α -carboethoxylation and stereoselective allylation to **7** and further addition of the Grignard reagent derived from 3-chloro-1-butene led to **8** as described previously (Scheme 1).^{7c} RCM reaction in **8** delivered the tricyclic product **9**. LAH reduction of **9** furnished the tricyclic diol **10a** to set-up the intended fragmentation reaction. Exposure of **10a** to methanesulfonyl chloride in the presence of base led to the isolation of a single product which was assigned the novel bridged structure **13** on the basis of incisive ¹H(COSY) and ¹³C NMR studies.⁹ Formation of **13** indicated that the Grob-type fragmentation in **10a** had indeed occurred through the mesylate **10b** to form the expected bicyclic neodolabellane-type product **11** (Scheme 1). However, in the reaction medium, **11** undergoes an extremely facile transannular carbonyl-ene cyclization to **12** through the participation of the exocyclic methylene group and further dehydration results in the observed product **13** (Scheme 1). It may be recalled that bridged cycloheptatriene derivatives similar to **13** have been encountered among sesquiterpenoid natural products,¹⁰ but no diterpenoid framework with this novel structural feature is known. Many variations of the Grob-type fragmentation in both **9** and **10a** were explored but it was not possible to isolate any 5,11-fused bicyclic product. Recourse was therefore taken to effect an oxy-Cope rearrangement¹¹ in a suitably crafted derivative of **5** to effect a four-carbon ring expansion.

To set up the precursor for the oxy-Cope process, hydroazulenone **5** was elaborated through sequential

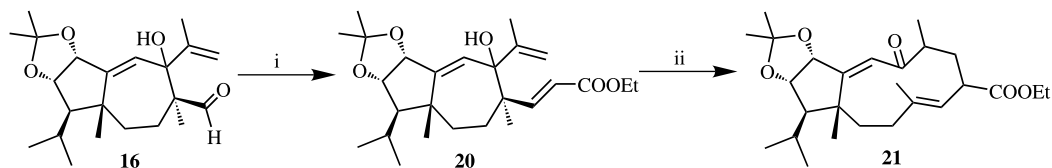
α -carboethoxylation and methylation to furnish **14** as a single diastereomer. Addition of the Grignard reagent prepared from 2-bromopropene to **14** was also stereoselective and furnished **15** (Scheme 2). The ester functionality in **15** was converted to the aldehyde **16** in two steps involving LAH reduction to the primary alcohol and reoxidation employing IBX. Wittig olefination of the aldehyde **16** afforded the divinyl carbinol **17**, the precursor for the oxy-Cope rearrangement (Scheme 2). Attempted, anionic oxy-Cope rearrangement on **17** employing KH or KHMDS or NaH as the base, with or without phase transfer catalysis and under different solvent regimes (THF, DME, dioxane) was singularly unsuccessful. However, thermal activation of **17** in boiling 1,2-dichlorobenzene induced the [3.3]-sigmatropic shift and oxy-Cope rearrangement products **18/19** were obtained as a mixture (1:1) of methyl epimers in modest yield (Scheme 2). While **18** and **19** could be separated and fully characterized, it was not possible to assign unambiguously stereochemistry to the newly generated secondary methyl group in either case despite many 2D NMR experiments. To demonstrate the generality of this oxy-Cope process, aldehyde **16** was subjected to a Horner–Wittig reaction to afford the (*E*)- α,β -unsaturated ester **20** as a single diastereomer. Thermal activation of **20**, under conditions identical to those employed for **17**, furnished the [3.3]-shift product **21**, quite fortuitously as a single diastereomer and in much better yield (Scheme 3). While the gross structure of **21** was readily forthcoming from ¹H and ¹³C NMR data, recourse to even high field 2D NMR (COSY, NOESY) studies was inadequate to deduce the stereostructure of this oxy-Cope rearrangement product.



Scheme 1. Reagents and conditions: (i) LDA, THF–HMPA, CNCOOEt, -78°C , 75%; (ii) NaH, THF, allyl bromide, rt, 95%; (iii) Mg, 3-chloro-1-butene, CeCl_3 , THF, 60%; (iv) cat. (10 mol%), benzene, Δ , 96%; (v) LAH, THF, Δ , 65–70%; (vi) NEt_3 , MsCl, DCM, 70%.



Scheme 2. Reagents and conditions: (i) LDA, THF–HMPA, CNCOOEt, -78°C , 75%; (ii) NaH, THF, MeI, rt, 95%; (iii) Mg, 2-bromopropene, THF, 0°C –rt, 80–85%; (iv) LAH, THF, rt, 98%; (v) IBX, toluene/DMSO (4:1), 85%; (vi) $\text{MePh}_3\text{P}^+\text{Br}^-$, KOtBu, THF, quant.; (vii) *o*-DCB, 180°C , 10 h, 35%.



Scheme 3. Reagents and conditions: (i) $(\text{EtO})_2\text{P}(\text{O})\text{CH}_2\text{COOEt}$, NaH, THF, rt, 2 h, 90%; (ii) *o*-DCB, 180°C , 6 h, 60%.

In summary, we have devised an oxy-Cope rearrangement based strategy to access the 5,11-fused bicyclic framework of novel neodolabellane diterpenes, replete with requisite stereochemical features at two key stereogenic centers and extensive functionalization, from a readily accessible hydroazulenic precursor.

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9. All new compounds reported here are racemic and were duly characterized on the basis of spectral (IR, ^1H and ^{13}C NMR) and analytical data. Selected spectral data: compound **10a**: ^1H NMR (300 MHz, CDCl_3) δ 5.66 (d, $J=1.5$ Hz, 1H), 5.64–5.61 (m, 1H), 5.34 (d, $J=10.8$ Hz, 1H), 4.86 (dd, $J=7.8$, 1.5 Hz, 1H), 4.38 (dd, $J=7.8$, 5.4 Hz, 1H), 3.51 (ABq, $J=10.8$ Hz, 2H), 2.54 (dt, $J=14.1$, 3.3 Hz, 1H), 2.31–2.03 (series of m, 4H), 1.82–1.68 (m, 2H), 1.48 (s, 3H), 1.38–1.31 (m, 2H), 1.36 (s, 3H), 1.12 (d, $J=7.2$ Hz, 3H), 1.06 (d, $J=6.6$ Hz, 3H), 1.05 (s, 3H), 0.99 (d, $J=6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 154.4, 130.2, 124.9, 123.1, 112.7, 83.0, 79.8, 72.9, 65.5, 62.2, 50.2, 46.4, 39.8, 36.7, 34.3, 30.1, 28.3, 27.9, 26.3, 23.3, 22.6, 18.3, 13.6; HRMS: calcd for $\text{C}_{23}\text{H}_{36}\text{O}_4\text{Na}$ ($\text{M}^+\text{+Na}$): 399.2511; Found: 399.2491; compound **13**: ^1H NMR (300 MHz, CDCl_3) δ 6.60 (s, 1H), 6.53–6.51 (m, 2H), 5.93 (d, $J=3$ Hz, 1H), 5.00 (d, $J=6.6$ Hz, 1H), 4.26 (dd as t $J=6.9$ Hz, 1H), 3.09 (d, $J=11.4$ Hz, 1H), 2.60 (t, $J=12.3$ Hz, 1H), 2.33 (dd, $J=12.3$, 7.2 Hz, 1H), 2.06 (dd, $J=14.7$, 6.9 Hz, 1H), 1.93 (s, 3H), 1.81–1.75 (m, 1H), 1.41 (s, 3H), 1.40 (merged signal, 1H), 1.38 (s, 3H), 1.31–1.25 (m, 1H), 1.23 (s, 3H), 1.09 (d, $J=6.6$ Hz, 3H), 1.02 (d, $J=6.6$ Hz, 3H), 0.88–0.83 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 147.1, 136.7, 133.6, 129.1, 128.9, 125.6, 120.9, 118.2, 111.8, 82.9, 79.9, 62.3, 49.6, 38.6, 37.5, 32.4, 28.2, 26.6, 26.4, 25.4, 21.2 (2C), 17.4; HRMS: calcd for $\text{C}_{23}\text{H}_{32}\text{O}_5\text{Na}$ ($\text{M}^+\text{+Na}$): 363.2300; Found: 363.2329; compound **17**: ^1H NMR (300 MHz, CDCl_3) δ 6.12 (dd, $J=18.0$, 10.5 Hz, 1H), 5.41 (d, $J=1.8$ Hz, 1H), 5.16 (br s, 1H), 5.08–4.96 (m, 3H), 4.80 (dd, $J=6.9$, 1.8 Hz, 1H), 4.37 (dd as t, $J=7.2$ Hz, 1H), 2.16 (dd as t, $J=13.2$ Hz, 1H), 1.95 (dd as t, $J=13.2$ Hz, 1H), 1.83 (s, 3H), 1.77–1.66 (m, 2H), 1.57–1.52 (m, 2H), 1.48 (s, 3H), 1.34 (s, 3H), 1.18 (s, 3H), 1.06 (d, $J=6.3$ Hz, 3H), 1.01 (d, $J=6.3$ Hz, 3H), 0.99 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 147.4, 147.3 (2C), 131.5, 115.1, 111.9, 110.6, 82.7, 82.4, 81.5, 63.2, 49.4, 45.5, 35.8, 30.1, 28.2, 27.3, 26.2, 24.1, 22.2, 21.9, 21.6, 20.0; HRMS: calcd for $\text{C}_{23}\text{H}_{36}\text{O}_3\text{Na}$ ($\text{M}^+\text{+Na}$): 383.2562; Found: 383.2572; compound **18**: ^1H NMR (300 MHz, CDCl_3) δ 6.32 (s, 1H), 5.09–5.05 (m, 1H), 4.68 (d, $J=6.3$ Hz, 1H), 4.46 (dd as t, $J=6.9$ Hz, 1H), 2.46–2.41 (m, 1H), 2.14–1.96 (m, 6H), 1.91–1.62 (m, 3H), 1.65 (s, 3H), 1.56 (s, 3H), 1.37 (s, 3H), 1.20–1.17 (m, 1H), 1.19 (s, 3H), 1.03 (d, $J=7.2$ Hz, 3H), 1.02 (d, $J=7.2$ Hz, 3H), 0.98 (d, $J=6.9$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 205.2, 156.8, 136.6, 128.6, 126.9, 111.5, 80.7, 51.6, 46.6, 33.8, 27.7, 27.3, 26.3, 26.1, 25.6 (br), 24.3, 23.9 (br), 17.4 (br); HRMS: calcd for $\text{C}_{23}\text{H}_{36}\text{O}_3\text{Na}$ ($\text{M}^+\text{+Na}$): 383.2562; Found: 383.2570; compound **19**: ^1H NMR (300 MHz, CDCl_3) δ 6.47 (s, 1H), 5.09 (dd, $J=11.1$, 6.0 Hz, 1H), 4.76 (d, $J=6.3$ Hz, 1H), 4.46 (dd as t, $J=6.9$ Hz, 1H), 2.91–2.84 (m, 1H), 2.24–2.03 (m, 4H), 1.89–1.68 (m, 4H), 1.62 (s, 3H), 1.52 (s, 3H), 1.39–1.33 (m, 1H), 1.38 (s, 3H), 1.20 (s, 3H), 1.11–1.05 (m, 1H), 1.06 (d, $J=6.3$ Hz, 6H), 1.01 (d, $J=6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 207.8, 159.1, 135.6, 128.6, 124.3, 111.9, 82.4, 81.9, 55.0, 52.4, 46.9, 33.8, 32.7, 28.2, 27.3, 26.6, 26.15, 25.2, 23.9, 23.7, 23.6, 21.6, 18.9; HRMS: calcd for $\text{C}_{23}\text{H}_{36}\text{O}_3\text{Na}$ ($\text{M}^+\text{+Na}$): 383.2562; Found: 383.2585; compound **20**: ^1H NMR (300 MHz, CDCl_3) δ 7.30 (d, $J=16.5$ Hz, 1H), 5.78 (d, $J=16.5$ Hz, 1H), 5.43 (d, $J=1.5$ Hz, 1H), 5.13 (br s, 1H), 5.02 (br s, 1H), 4.82 (dd, $J=6.9$, 1.8 Hz, 1H), 4.39 (dd as t, $J=6.9$

Hz, 1H), 4.18 (q, $J=6.9$ Hz, 2H), 2.14 (dd as t, $J=13.2$ Hz, 1H), 1.96 (dd as t, $J=13.2$ Hz, 1H), 1.83–1.69 (m, 2H), 1.77 (s, 3H), 1.59–1.52 (m, 2H), 1.48 (s, 3H), 1.34 (s, 3H), 1.28 (t, $J=6.9$ Hz, 3H), 1.23 (s, 3H), 1.07 (d, $J=6.6$ Hz, 3H), 1.01 (d, $J=6.6$ Hz, 3H), 0.98 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 167.2, 157.6, 146.8, 146.7, 131.1, 117.1, 115.7, 112.0, 82.6, 82.3, 81.4, 63.2, 60.2, 49.6, 45.8, 35.7, 30.0, 28.1, 27.3, 26.1, 24.1, 22.1, 21.7, 21.4, 19.8, 14.3; HRMS: calcd for $\text{C}_{26}\text{H}_{40}\text{O}_5\text{Na}$ ($\text{M}^+ + \text{Na}$): 455.2773; Found: 455.2774; compound **21**: ^1H NMR (300 MHz, CDCl_3) δ 6.24 (s, 1H), 5.74 (d, $J=10.8$ Hz, 1H), 4.67 (d, $J=6.3$ Hz, 1H), 4.46 (t, $J=6.6$ Hz, 1H), 4.11 (q, $J=6.9$ Hz, 2H), 3.27–3.20 (m, 1H), 2.50–2.30 (m, 2H), 2.00–1.96 (m, 1H), 1.88–1.77 (m, 3H), 1.74 (d, $J=9.0$ Hz, 3H), 1.57 (s, 3H), 1.37 (s, 3H), 1.26–1.22 (m, 8H), 1.10–1.06 (m,

1H), 1.03 (d, $J=6.6$ Hz, 3H), 1.03 (s, 3H), 1.02 (d, $J=6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 205.5, 173.9, 157.2, 138.1, 128.8, 125.3, 111.5, 60.5, 51.2 (br), 45.6, 45.5, 37.6, 29.7, 27.5 (br), 26.2, 25.4 (br), 24.0 (br), 20.7 (br), 19.0, 14.2; HRMS: calcd for $\text{C}_{26}\text{H}_{40}\text{O}_5\text{Na}$ ($\text{M}^+ + \text{Na}$): 455.2773; Found: 455.2792.

10. Spiniferin sesquiterpenes having bicyclo[4.4.1^{1,6}]undecane skeleton incorporate a bridged cycloheptatriene moiety. Isolation: (a) Cimino, G.; De Stefano, S.; Minale, L.; Trivellone, E. *Tetrahedron Lett.* **1975**, 26, 3727; (b) Cimino, G.; De Stefano, S.; Minale, L.; Trivellone, E. *Experientia* **1978**, 34, 1425. Synthesis: Marshall, J. A.; Conrow, R. E. *J. Am. Chem. Soc.* **1980**, 102, 4274.
11. (a) Paquette, L. A. *Tetrahedron* **1997**, 53, 13971; (b) Wilson, S. R. *Org. React.* **1993**, 43, 93.